



22-ID-05

Committee: Infectious Disease

Title: Update to the Standardized Case Definition and National Notification for Candida auris

☑Check this box if this position statement is an update to an existing standardized surveillance case definition and include the most recent position statement number here: 18-ID-05 .

Synopsis: This position statement updates the *Candida auris* case definition by removing presumptive laboratory criteria and probable and suspect case classifications and by recommending *C. auris* screening cases be made nationally notifiable. Updates include new information on transmissibility and case counts.

I. Statement of the Problem

Candida auris is an emerging multidrug-resistant yeast that can colonize the skin and cause invasive infections. It can spread readily between patients in healthcare facilities, causing numerous outbreaks that have been difficult to control. Containment of *C. auris* spread largely depends on timely detection and implementation of appropriate infection prevention and control measures (1).

Individuals colonized on their skin can be identified through screening tests; they can shed into the environment, thereby presenting similar transmission risks and requiring the same infection control precautions as individuals with *C. auris* identified in clinical specimens. Screening detects outbreaks earlier than relying solely on passive surveillance through clinical specimens; in several large outbreaks, a majority of incident cases were identified through screening tests (2, 3). Currently, however, only cases identified through clinical specimens are nationally notifiable. Incomplete reporting of cases could lead to delayed identification and outbreak response and prevent jurisdictions from understanding the full burden of *C. auris* needed to guide public health action.

Since 2018, laboratory capability to detect *C. auris* has greatly improved. Misidentification of *C. auris* (particularly as *C. haemulonii*) is much less common than it was previously, and confirmatory testing is now widely available; only three probable or suspect cases with unconfirmed laboratory evidence have been reported nationally since 2018, and none were reported in 2021. The presumptive laboratory evidence criteria (detection of *C. haemulonii* with a laboratory method unable to detect *C. auris*, without confirmatory testing), and related probable and suspect case classifications, are no longer useful and add unnecessary complexity to reporting.

A case definition, adopted in 2017 and updated in 2018, allowed for standardized tracking of *C. auris* cases (4). This position statement updates the case definition to reflect improved laboratory capability to identify *C. auris* and highlights the importance of reporting screening cases nationally.

II. Background and Justification

C. auris was classified as an urgent threat in the 2019 Antibiotic Resistance Threats Report (5). Some strains are resistant to all three major classes of antifungals, severely limiting treatment options (6). It can cause invasive infections and is associated with 30-72% crude in-hospital mortality (7). *C. auris* can colonize patients' skin and other body sites for prolonged periods, and colonization poses a risk both for invasive infection and transmission. *C. auris* persists in the healthcare environment for weeks, and certain routinely-used disinfectants in healthcare settings are not effective against the organism (8, 9). *C. auris* can spread rapidly within healthcare facilities, especially in high-acuity long-term care settings, colonizing large proportions of patients (10). Outbreaks of *C. auris* have proven very difficult to control, requiring intensive public health and facility-level interventions (3, 11).

As of January 1, 2022, more than 3,200 cases have been identified through clinical specimens and more than 7,300 through screening specimens across 29 states. The extent of *C. auris* transmission is variable across the



Council of State and Territorial Epidemiologists

United States, and there is greater opportunity to control spread in less affected areas and prevent widespread transmission across the country. Reporting of both clinical and screening cases is critical as public health and facility responses generally do not differ by case type. Further, colonization can lead to clinical infections; in one outbreak, 4% of patients colonized with *C. auris* developed bloodstream infections (12). Timely identification of colonization and subsequent response efforts can help prevent additional transmission and related morbidity and mortality among high-risk patients. This position statement makes screening cases nationally notifiable, which in turn would facilitate reporting changes at the state and local level. This would enable prompt detection, investigation, and response, including the coordination of transfers between facilities and jurisdictions, and other measures critical to containing the spread of *C. auris*.

C. auris cases identified through screening tests only indicate colonization; however, clinical cases (such as those indicated by urine and respiratory specimens) can represent colonization if no evidence of clinical infection exists. Because there is overlap between case types, this can cause unnecessary confusion during reporting. We propose changing the nomenclature to reflect how cases are identified (i.e., through clinical or screening specimens), rather than the differentiation between infection and colonization. Therefore, patients identified through screening tests (e.g., axilla/groin swab) would be classified as screening cases rather than colonization/screening cases.

C. auris has historically been misidentified by some laboratory methods, making detection and therefore control of C. auris challenging; however, clinical and public health laboratory capability to identify and confirm C. auris has expanded greatly. Therefore, the current presumptive laboratory evidence criteria, which includes the detection of C. haemulonii with a laboratory method unable to detect C. auris, is no longer as relevant. This case definition removes this criterion, as well as the probable clinical case, suspect clinical case, and probable screening case classifications that rely on presumptive laboratory evidence.

III. Statement of the desired action(s) to be taken

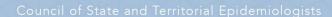
CSTE recommends the following actions:

- 1. Implement a standardized surveillance case definition for Candida auris.
 - A. Utilize standard sources (e.g., reporting*) for case ascertainment for *Candida auris*. Surveillance for *Candida auris* should use the recommended sources of data to the extent of coverage presented in Section V.
 - B. Utilize standardized criteria for case ascertainment for *Candida auris* presented in Section VI and Table VI in Technical Supplement.
 - C. Utilize standardized criteria for case classification for *Candida auris* presented in Section VII and Table VII in Technical Supplement.

2.	Utilize standardized criteria for case ascertainment and classification (based on Sections VI and VII and
	Technical Supplement) for Candida auris and update Candida auris on the Nationally Notifiable Condition
	List.
	☐ Immediately notifiable, extremely urgent (within 4 hours)
	☐ Immediately notifiable, urgent (within 24 hours)
	☒ Routinely notifiable, clinical and screening

☐ No longer notifiable

3. CSTE recommends that all States and Territories enact laws (statute or rule/regulation as appropriate) to make this disease or condition reportable in their jurisdiction. Jurisdictions (e.g., States and Territories) conducting surveillance (according to these methods) should submit case notifications** to CDC.





4. Expectations for Message Mapping Guide (MMG) development for a newly notifiable condition: the National Notifiable Diseases Surveillance System (NNDSS) is transitioning to HL7-based messages for case notifications; the specifications for these messages are presented in MMGs. When CSTE recommends a new condition be made nationally notifiable, CDC must obtain Office of Management and Budget Paperwork Reduction Act (OMB PRA) approval prior to accepting case notifications for the new condition. Under anticipated timelines, notification using the Generic V2 MMG would support transmission of the basic demographic and epidemiologic information common to all cases and could begin with the new MMWR year following the CSTE annual conference. Input from CDC programs and CSTE would prioritize development of a disease-specific MMG for the new condition among other conditions waiting for MMGs.

5.	CDC should publish data on clinical and screening cases of Candida auris as appropriate (see Section IX)
	CSTE recommends the following case statuses be included in the CDC Print Criteria:
	⊠ Confirmed
	□Probable
	□Suspect
	□Unknown

6. CSTE recommends that all jurisdictions (e.g., States, Localities, or Territories) with legal authority to conduct public health surveillance follow the recommended methods outlined in this recommendation and in the accompanying standardized surveillance position statement.

IV. Goals of Surveillance

To assess the temporal, geographic, and demographic burden of *C. auris* in the United States to facilitate response, prevention, and containment.

V. Methods for Surveillance: Surveillance for Candida auris should use the recommended sources of data and the extent of coverage listed in Table V.

The primary sources of data are microbiology laboratory results. State, territorial, local, and tribal (STLT) public health agencies may also utilize data obtained from clinician reporting or other entities to ascertain cases. Laboratories that identify an organism that might represent *C. auris* in a specimen or isolate using a test that could misidentify it should obtain confirmatory testing (13).

Table V. Recommended sources of data and extent of coverage for ascertainment of cases of *Candida auris*.

	Coverage	
Source of data for case ascertainment	Population-wide	Sentinel sites
Clinician reporting	X	
Laboratory reporting	X	
Reporting by other entities, specify: acute care	X	
hospitals, long-term care facilities, and outpatient		
settings		
Death certificates		
Hospital discharge or outpatient records		

^{*}Reporting: process of a healthcare provider or other entity submitting a report (case information) of a condition under public health surveillance to local, state, or territorial public health.

^{**}Notification: process of a local or state public health authority submitting a report (case information) of a condition on the Nationally Notifiable Conditions List to CDC.





Data from electronic medical records	X	
Telephone survey		
School-based survey		
Other, specify: N/A		

VI. Criteria for case ascertainment

A. Narrative: A description of suggested criteria for case ascertainment of a specific condition.

A1. Clinical Criteria for Reporting

N/A

A2. Laboratory Criteria for Reporting

Report any patient or laboratory finding to public health authorities that meets the following criterion:

• Detection of *C. auris* in a specimen using either culture or a validated culture-independent test (e.g., nucleic acid amplification test [NAAT])

A3. Epidemiologic Linkage Criteria for Reporting

N/A

B. Disease-specific data elements to be included in the initial report

In addition to patient demographics, the following disease-specific data elements are expected to be included in all reports to public health agencies:

- Specimen source
- Indication for testing (i.e., clinical or screening)
- Collection date
- Results

VII. Case Definition for Case Classification

A. Narrative: Description of criteria to determine how a case should be classified.

A1. Clinical Criteria

N/A

A2. Laboratory Criteria

Confirmatory laboratory evidence:

- Detection of *C. auris* in a specimen from a swab obtained for the purpose of colonization screening using either culture or validated culture-independent test (e.g., nucleic acid amplification test [NAAT]), OR
- Detection of *C. auris* in a clinical specimen obtained during the normal course of care for diagnostic or treatment purposes using either culture or a validated culture-independent test (e.g., NAAT)

Presumptive laboratory evidence:

N/A





Supportive laboratory evidence: N/A

Note: The categorical labels used here to stratify laboratory evidence are intended to support the standardization of case classifications for public health surveillance. The categorical labels should not be used to interpret the utility or validity of any laboratory test methodology.

A3. Epidemiologic Linkage

N/A

A4. Case Classifications

Confirmed:

- <u>Candida auris case</u>, <u>screening</u>: Person with confirmatory laboratory evidence from a swab collected for the purpose of screening for *C. auris* colonization regardless of site swabbed.*
- <u>Candida auris</u> case, clinical: Person with confirmatory laboratory evidence from a clinical specimen collected for the purpose of diagnosing or treating disease in the normal course of care.**

Probable: N/A

Suspect: N/A

- * Typical screening specimen sites are skin (e.g., axilla, groin), nares, rectum, or other external body sites. Swabs collected from wound or draining ear as part of clinical care are considered clinical specimens.‡
- ** This includes specimens from sites reflecting invasive infection (e.g., blood, cerebrospinal fluid) and specimens from non-invasive sites such as wounds, urine, and the respiratory tract, where presence of C. auris may simply represent colonization and not true infection. This does not include swabs collected for screening purposes (see Candida auris case, screening).
- [‡]Because it can be difficult to differentiate screening specimens from clinical specimens based on microbiology records, any swabs except wound swabs or draining ear swabs can be assumed to be for screening unless specifically noted otherwise. Laboratories do not need to change their practice; public health wants to identify all C. auris whether from screening or clinical specimens.

B. Criteria to distinguish a new case of this disease or condition from reports or notifications which should not be enumerated as a new case for surveillance

A patient who is colonized or infected with *C. auris* is considered colonized indefinitely. The following provides guidance for health departments to distinguish a new case for patients who test positive for *C. auris* in either a screening swab (i.e., screening case) or in a clinical specimen (i.e., clinical case).

- For screening cases, count patient only once as a screening case; do not count if patient has been
 previously identified as a clinical or screening case. A person with a screening case can be later
 categorized as a clinical case (e.g., patient with positive screening swab who later develops bloodstream
 infection would be counted in both categories).
- For clinical cases, count patient only once as a clinical case, even if the patient has already been counted separately as a screening case. A person with a clinical case should not be counted as a screening case thereafter because all clinical cases are considered to also be colonized with *C. auris* (e.g., patient with clinical *C. auris* specimen who later has positive screening swab is not counted as a screening case).

VIII. Period of Surveillance

Ongoing





IX. Data sharing/release and print criteria

ends the following case statuses* be included in the 'case' count released outside of the
gency:
⊠Confirmed
□Probable
□Suspect
□Unknown

Jurisdictions (e.g., States and Territories) conducting surveillance under this case definition can voluntarily submit de-identified case information to CDC, if requested and in a mutually agreed upon format.

Production of national data summaries and national data re-release for non-NNCs:

- Prior to release of national data summaries CDC should follow the CDC/ATSDR Policy on Releasing & Sharing Data, issued on April 16, 2003 and referenced in 11-SI-01 and custodians of such data should consult the CDC-CSTE Intergovernmental Data Release Guidelines Working Group report (www.cste2.org/webpdfs/drgwgreport.pdf) which contains data release guidelines and procedures for CDC programs re-releasing state, local, or territorial-provided data.
- CDC programs have a responsibility, in collaboration with states, localities, and territories, to ensure that CDC program-specific data re-release procedures meet the needs of those responsible for protecting data in the states and territories.

X. Revision History

Previous PS ID	Section of Document	Revision Description
18-ID-05	VI. Criteria for case ascertainment	Removed detection of an organism that commonly represents a <i>C. auris</i> misidentification in a specimen by culture under laboratory criteria. Added disease-specific elements to criteria for case ascertainment.
18-ID-05	VII. Case Definition for Case Classification	Removed probable and suspect status for both clinical and screening cases. Removed associated definitions for presumptive laboratory evidence and epidemiologic linkages for the case classifications being removed.
18-ID-05	IX. Data Sharing	Removed recommendation that probable cases be included in the case count criteria.
18-ID-05	NNC Recommendation Statement	Recommended that confirmed screening cases be included in CDC print criteria by jurisdiction.
18-ID-05	Appendix I	Removed Appendix I pertinent to presumptive laboratory criteria for case classification. Included CDC website as citation in references.
18-ID-05	VII. Case classification	December 2018: Authors made non-substantive changes to add clarity for implementation.
17-ID-03	II. Background and Justification	Updated with new information about transmissibility of <i>C. auris</i> and case counts in the U.S.
17-ID-03	III. Statement of desired actions to be taken	Table III, removed the following sources of data: death certificates, hospital discharge or outpatient records, extracts from, electronic medical records. For coverage continue population-wide, removed sentinel sites.

^{*} Which case statuses are included in the case counts constitute the "print criteria."





17-ID-03	VII. Case definition, laboratory criteria	Revised to reflect updates in laboratory test performance characteristics, include CIDT in addition to culture, refer to Appendix 1 instead of text within position statement for details of misidentifications. Changed label from supportive to presumptive laboratory criteria. Added under presumptive lab criteria that the isolate/specimen has not yet undergone further testing. Added clarifying note: When additional test results are available, case re-classification may occur, including making this a non-case.
17-ID-03	VII. Case definition, epidemiologic linkage criteria	Added epidemiologic linkage to patients with presumptive laboratory evidence (in addition to confirmatory); clarify that no overlapping time- period is required, add time-frame (12 months) for epidemiological linkage, add overnight stay in healthcare facility overseas in previous one year in foreign country with documented <i>C. auris</i> transmission.
17-ID-03	VII. Case classification	Changed "screening" to "colonization/screening" Added probable colonization/screening case classification. Clarified that swabs from wounds or draining ears are considered clinical.
17-ID-03	IX. Data	Added CDC Print Criteria
17-ID-03	XI. References	Added/updated references
17-ID-03	NNC Recommendation Statement	Recommends adding <i>Candida auris</i> (clinical) to the Nationally Notifiable Condition List as routinely notifiable (only clinical). Statement on message mapping guide: CSTE recommends that a working group be established that includes CSTE and CDC members.
17-ID-03	Appendix 1	Added appendix 1 that describes <i>C. auris</i> identification methods including common misidentifications
N/A	17-ID-03	Creates a standardized case definition for <i>Candida auris</i> causing clinical infection and colonization.

XI. References

- Caceres DH, Forsberg K, Welsh RM, et al. *Candida auris*: A Review of Recommendations for Detection and Control in Healthcare Settings. J Fungi (Basel). 2019;5(4):111. Published 2019 Nov 28. doi:10.3390/jof5040111
- 2. Zhu Y, O'Brien B, Leach L, et al. Laboratory Analysis of an Outbreak of Candida auris in New York from 2016 to 2018: Impact and Lessons Learned. *J Clin Microbiol*. 2020;58(4):e01503-19. Published 2020 Mar 25. doi:10.1128/JCM.01503-19
- 3. Karmarkar EN, O'Donnell K, Prestel C, et al. Rapid Assessment and Containment of *Candida auris* Transmission in Postacute Care Settings-Orange County, California, 2019. *Ann Intern Med*. 2021;174(11):1554-1562. doi:10.7326/M21-2013
- 4. CSTE. Standardized Case Definition for *Candida auris* clinical and colonization/screening cases and National Notification of *C. auris* case, clinical. https://cdn.ymaws.com/www.cste.org/resource/resmgr/2018 position statements/18-ID-05.pdf
- 5. CDC. Antibiotic Resistance Threats in the United States, 2019. Atlanta, GA: U.S. Department of Health and Human Services, CDC; 2019 http://www.cdc.gov/drugresistance/Biggest-Threats.html
- 6. Lockhart SR, Etienne KA, Vallabhaneni S, et al. Simultaneous Emergence of Multidrug-Resistant *Candida auris* on 3 Continents Confirmed by Whole-Genome Sequencing and Epidemiological Analyses [published correction appears in Clin Infect Dis. 2018 Aug 31;67(6):987]. *Clin Infect Dis*. 2017;64(2):134-140. doi:10.1093/cid/ciw691





- 7. Cortegiani A, Misseri G, Fasciana T, Giammanco A, Giarratano A, Chowdhary A. Epidemiology, clinical characteristics, resistance, and treatment of infections by *Candida auris*. *J Intensive Care*. 2018;6:69. Published 2018 Oct 29. doi:10.1186/s40560-018-0342-4
- 8. Cadnum JL, Shaikh AA, Piedrahita CT, et al. Effectiveness of Disinfectants Against *Candida auris* and Other Candida Species. *Infect Control Hosp Epidemiol*. 2017;38(10):1240-1243. doi:10.1017/ice.2017.162
- 9. Sexton DJ, Bentz ML, Welsh RM, et al. Positive Correlation Between *Candida auris* Skin-Colonization Burden and Environmental Contamination at a Ventilator-Capable Skilled Nursing Facility in Chicago. *Clin Infect Dis.* 2021;73(7):1142-1148. doi:10.1093/cid/ciab327
- 10. Pacilli M, Kerins JL, Clegg WJ, et al. Regional Emergence of *Candida auris* in Chicago and Lessons Learned From Intensive Follow-up at 1 Ventilator-Capable Skilled Nursing Facility. *Clin Infect Dis*. 2020;71(11):e718-e725. doi:10.1093/cid/ciaa435
- 11. Adams E, Quinn M, Tsay S, et al. *Candida auris* in Healthcare Facilities, New York, USA, 2013-2017. *Emerg Infect Dis.* 2018;24(10):1816-1824. doi:10.3201/eid2410.180649
- Southwick K, Adams EH, Greenko J, et al. 2039. New York State 2016–2018: Progression from Candida auris Colonization to Bloodstream Infection. Open Forum Infect Dis. 2018;5(Suppl 1):S594-S595. Published 2018 Nov 26. doi:10.1093/ofid/ofy210.1695
- 13. Identification of *Candida auris*. Centers for Disease Control and Prevention. Updated May 29, 2020. Accessed February 18, 2022. https://www.cdc.gov/fungal/candida-auris/identification.html

XII. Coordination

Subject Matter Expert (SME) Consultants:

PRIMARY SME

(1) Meghan Lyman MD
Medical Epidemiologist
Centers for Disease Control and Prevention
404-639-4241
yeo4@cdc.gov

ADDITIONAL SME(s)

(1) Kaitlin Forsberg, MPH
Epidemiologist
Centers for Disease Control and Prevention
404-718-3575
Inv6@cdc.gov

(2) Brendan Jackson MD, MPH
Medical Epidemiologist
Centers for Disease Control and Prevention
404-639-4241
brjackson1@cdc.gov

Agency for Response

(1) Centers for Disease Control and Prevention Rochelle Walensky, MD
Director
1600 Clifton Road NE
Atlanta, GA 30329
404-639-7000
Aux7@cdc.gov





Agencies for Information:

- (1) Infectious Disease Society of America
 Daniel P McQuillen, MD, FIDSA
 President
 4040 Wilson Boulevard
 Suite 300
 Arlington, VA 22203
 703-229-0200
- (2) Society for Healthcare Epidemiology of America
 Kristy Weinshel, MBA, CAE
 Executive Director
 4040 Wilson Boulevard
 Suite 300
 Arlington, VA 22203
 703-684-1006
 kweinshel@shea-online.org
- (3) Association for Professionals in Infection Control Devin Jopp, EdD, MS
 Chief Executive Officer
 1400 Crystal Drive
 Suite 900
 Arlington, VA 22202
 202-454-2628
 djopp@apic.org
- (4) Association for Public Health Laboratories Scott Becker, MS Executive Director 8515 Georgia Ave #700 Silver Spring, MD 20910 Scott.Becker@aphl.org

XIII. Author Information

Submitting and Presenting Author:

(1) Sam Horwich-Scholefield MPH, CIC
Antimicrobial Resistance Epidemiologist
California Department of Public Health
850 Marina Bay Parkway
Building E, First Floor
Richmond, CA 94804
510-412-6060
sam.horwich-scholefield@cdph.ca.gov

Co-Author(s):

- (1) Active Member
 Kelly Walblay MPH
 Epidemiologist IV Healthcare Settings
 Chicago Department of Public Health
 1340 S Damen, 4th Floor
 Chicago, IL 60608
 312-746-8223
 kelly.walblay@cityofchicago.org
- (2) Active Member
 Tisha Mitsunaga DrPH, ScM
 Research Scientist Supervisor
 California Department of Public Health
 850 Marina Bay Parkway
 Building E, First Floor
 Richmond, CA 94804
 510-412-6060
 tisha.mitsunaga@cdph.ca.gov

Additional co-authors are included in Appendix 1.



Council of State and Territorial Epidemiologists

Technical Supplement

Table VI. Table of criteria to determine whether a case should be reported to public health authorities.

Criterion	Candida auris
Clinical Criteria for Reporting	
N/A	
Laboratory Criteria for Reporting	
Detection of <i>C. auris</i> in a specimen using either culture or a validated	S
culture-independent test (e.g., nucleic acid amplification test [NAAT])	
Epidemiological Linkage Criteria for Reporting	
N/A	

Notes:

S = This criterion alone is SUFFICIENT to report a case.

Table VII. Classification Table: Criteria for defining a case of Candida auris.

Criterion	Confirmed – Screening Case	Confirmed – Clinical Case
Clinical Evidence		
N/A		
Laboratory Evidence		
Detection of <i>C. auris</i> in a specimen from a swab obtained for the purpose of colonization screening using either culture or validated culture-independent test (e.g., nucleic acid amplification test [NAAT]).	S	
Detection of <i>C. auris</i> in a clinical specimen obtained during the normal course of care for diagnostic or treatment purposes using either culture or a validated culture-independent test (e.g., NAAT).		S
Epidemiologic Linkage Evidence		
N/A		
Criteria to distinguish a new case:		
For screening cases, count patient only once as a screening case; do not count if patient has been	N	
previously identified as a clinical or screening case.		
For clinical cases, count patient only once as a clinical case, even if the patient has already been counted separately as a screening case.		N

Notes

S = This criterion alone is SUFFICIENT to classify a case.

N = All "N" criteria in the same column are NECESSARY to classify a case.



Council of State and Territorial Epidemiologists

Appendix 1. Additional Co-Authors

Numbering continues from Section XIII of the accompanying position statement.

(3) Sandeep K. Bhaurla MPH, CIC

Epidemiologist
Los Angeles County Department of Public Health
313 N. Figueroa St, Room 212
Los Angeles, CA, 90012
213-240-741
sbhaurla@ph.lacounty.gov
Active Member

(4) Joseph Gerth, MPH

Surveillance Epidemiologist, Division of Surveillance, Analytics, and Informatics
Massachusetts Department of Public Health
305 South Street
Boston, MA 02130
781-223-6757
Joseph.m.gerth@mass.gov
Active Member

(5) Diana Holden, MPH

Antimicrobial Resistance Epidemiologist Supervisor California Department of Public Health 850 Marina Bay Parkway Building E, First Floor Richmond, CA 94804 (510) 340-1357 Diana.Holden@cdph.ca.gov Active Member

(6) Gabriel K. Innes, VMD, PhD, CIC, DACVPM

Antimicrobial Resistance Surveillance Coordinator Infection Control, Healthcare, & Environmental Epi Infectious and Zoonotic Disease Program Communicable Disease Service
New Jersey Department of Health
P.O. Box 369
Trenton, NJ 08625-0369
Gabriel.innes@doh.nj.gov
(609) 913-5232
Active Member

(7) Meghan Maloney, MPH

Epidemiologist 4
Healthcare-Associated Infections/ Antimicrobial Resistance Program
Connecticut Department of Public Health
410 Capitol Ave., MS# 11EPI
PO Box 340308
Hartford, CT 06134-0308
860-509-7752
meghan.maloney@ct.gov
Active Member

(8) Julie Paoline, MA, CPHA, CIC, FAPIC

Public Health Specialist Healthcare-associated Infection Prevention/Antimicrobial Stewardship Pennsylvania Department of Health 625 Forster St. Harrisburg, PA 17120 717-547-3498 <u>c-jpaoline@pa.gov</u> Active Member

(9) Judi Sedivy MPH, BSN, RN CIC

Epidemiology Research Associate
Healthcare-associated Infection Prevention/
Antimicrobial Stewardship
Pennsylvania Department of Health
233 West Otterman Street
Greensburg, PA 15601
724-830-2701
jsedivy@pa.gov
Active Member

(10) Adrienne Sherman, MPH

Healthcare Associated Infections and Antimicrobial Resistance Epidemiologist Infection Control, Healthcare, & Environmental Epidemiology Infectious and Zoonotic Disease Program Communicable Disease Service New Jersey Department of Health P.O. Box 369
Trenton, NJ 08625-0369
Phone: 609-422-7051
Adrienne.Sherman@doh.nj.gov
Active Member

(11) Carolyn Stover, MPH, CPH

Epidemiologist, Healthcare Associated Infections and Antimicrobial Resistance Program
Tennessee Department of Health
710 James Robertson Parkway
Nashville, TN 37243
615-253-1273
Carolyn.Stover@tn.gov
Active Member

(12) Laura Tourdot, MPH

Epidemiologist Senior Minnesota Department of Health 625 Robert Street North Saint Paul, MN 55155 651-201-4361 laura.tourdot@state.mn.us Active Member

(13) Erica Washington, MPH, CPH, CIC, CPHQ, FAPIC

Healthcare-Associated Infections & Antibiotic Resistance Program Coordinator Public Health Epidemiologist Manager Infectious Disease Epidemiology Section Louisiana Department of Public Health 1450 Poydras Street, Ste. 1641 New Orleans, LA 70112 (504) 568-8319 <u>Erica.Washington@LA.GOV</u>

Appendix: 22-ID-05